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(54) Title: MUTATED GENE CODING FOR A LAT PROTEIN AND THE BIOLOGICAL APPLICATIONS THEREOF

(57) Abstract: The present invention relates to a mutated gene coding for a mutant LAT protein leading to an exaggerated T_H2 differentiation. The invention relates to biological structures containing said mutant, particularly, non-human LAT gene mutated animals, plasmids, chromosomal DNA's, embryos comprising said mutated gene, and applications thereof. The invention further relates to screening method for drug useful for treatment against asthma and allergy. Otherwise, the invention relates to method for producing IgE antibodies.

**Mutated gene coding for a LAT protein
and the biological applications thereof.**

The present invention relates to a mutated gene coding for two mutant LAT proteins leading to an exaggerated T_H2 cell differentiation. The invention relates to biological structures containing said mutant, particularly, non-human
5 LAT gene mutated animals, cell cultures, plasmids, chromosomal DNAs, embryos comprising said mutated gene, and applications thereof. The invention further relates to screening methods for drug useful for treatment against asthma, allergy and any pathological immune responses
10 involving T_H2 cells. The invention also relates to method for producing IgE antibodies.

Background Art

A key event in the pathogenesis is the production of
15 antibodies of the IgE class. Hypergammaglobulinemia E results from loss of immunoregulation. More specifically, T lymphocyte abnormalities have been reported in a number of pathologic hyper IgE conditions and are the object of much research aiming at developing pharmaceutical compounds that
20 will prevent atopic allergy and asthma.

TCR recognize peptide fragments bound to major histocompatibility complex (MHC) molecules and relay this information to the interior of the T cell via adapter
25 proteins. One of these, the adapter LAT (Linker for Activation of T cells), coordinates the assembly of signaling complexes through multiple tyrosine residues within its intracytoplasmic segment. Upon TCR-induced phosphorylation, each of these tyrosine residues manifests
30 some specialization in the signaling proteins it recruits. Studies on cell lines showed that mutation of tyrosine 136 (Y136) selectively eliminates binding of phospholipase $C\gamma 1$ (PLC- $\gamma 1$) whereas the simultaneous mutation of Y175, Y195 and

Y235 results in loss of binding of downstream adapters Gads and Grb-2 (Lin and Weiss, 2001; Samelson et al, 1999 ; Zhang et al, 2000). Studies of LAT "knock in" mutant mice presenting the mutation of the four distal tyrosine residues of LAT in phenylalanine, called 4YF mice, showed that the murine T cell development was completely blocked (Sommers et al, 2001). Hence, their thymocyte development was arrested at the immature CD4⁻ CD8⁻ stage and no mature T cells were present.

10

The present invention now provides genetic evidence that LAT exerts an unanticipated and surprising inhibitory function on the differentiation of CD4 helper T (T_H) cells into T_H2 cells.

15

Mice homozygous for the mutation of a single LAT tyrosine (LAT Y136F) results in mice that show a precocious and spontaneous accumulation of polyclonal T_H2 cells, which chronically produce large amounts of interleukins 4, 5, 10 and 13. This exaggerated T_H2 differentiation leads in turn to tissue eosinophilia and to the maturation of massive numbers of plasma cells secreting IgE and IgG1 antibodies (see Figure 1). Thus, in addition to known positive signaling, LAT also appears essential for establishing inhibitory signals that control T cell homeostasis.

Mice for the composite mutation of the three distal LAT tyrosines (LAT Y175F+Y195F+Y235F) prevents the development of T cells expressing alpha/beta T cell receptor. However, it allows the development of T cells expressing gamma/delta T cell receptors, and their accumulation in the periphery (see figure 9). These polyclonal gamma/delta T cells chronically produce large amounts of interleukins 4, 5, 10 and 13 (i.e. they present blatant TH2 phenotype). This exaggerated T_H2-type differentiation of gamma/delta T cells leads in turn to the maturation of massive numbers of plasma cells secreting IgE and IgG1 antibodies (see Figures 10 and

11).

Description of drawings

Figure 1 is a diagram disclosing the immune system
5 development of mutant mice.

Figure 2 illustrates the LAT Y136F knock-in strategy:

(1) : the partial restriction map of the wild-type LAT
gene.

(2) : the targeting vector used for the introduction of
10 the LAT Y136F mutation.

(3) : the structure of the targeted allele following
homologous recombination.

(4) : the final structure of the targeted allele after
removal of the neo^r gene via Cre-mediated recombination.

15 Figure 3 illustrates the aberrant growth of lymphoid organs
in the mice : thymus (A), spleen (B) and lymph nodes (C).

Figure 4 relates to constitutive type-2 cytokine production
in CD4 T cells freshly isolated from LAT^{Y136F} peripheral
lymphoid organs.

20 Figure 5 relates to a phenotypic analysis of T cells from
wild-type and LAT^{Y136F} mice.

Figure 6 illustrates eosinophilia in 6 weeks old LAT^{Y136F}
lymphoid organs.

A: Dot plot panel showing the gate selected for the
25 analysis described in panel B and for the sorted cells
picture in panel C.

B: Single color histograms of gated cells labelled with
antibodies characterizing eosinophils.

C: Hematoxylin and eosin staining of sorted cells.

30 Figure 7 illustrates the hyperactivity of B lymphocytes:
massive serum levels of IgE and IgG1 antibodies in
unimmunized LAT^{Y136F} mice.

Figure 8 illustrates the LAT Y175F+Y195F+Y235F knock in
strategy:

35 (1) : the partial restriction map of the wild-type LAT
gene.

(2) : the targeting vector used for the introduction of

the LAT Y175F, Y195F and Y235F mutation.

(3) : the structure of the targeted allele following homologous recombination.

(4) : the final structure of the targeted allele after
5 removal of the neo^r gene via Cre-mediated recombination.

Figure 9 relates to a phenotypic analysis of the gamma/delta T cells developed in large numbers in the LAT Y175F+Y195F+Y235F mutant in the mere absence of alpha/beta T cells.

10 Figure 10 illustrates the T_H2-type cytokines that are spontaneously produced by the gamma/delta T cells present in LAT Y175F+Y195F+Y235F mutant mice (lane 3) and compare them the T_H2-type cytokines that are spontaneously produced by the alpha/beta T cells developed in the LAT Y136F mutant (lane
15 1).

Figure 11 illustrates the hyperactivity of B lymphocytes and the massive amounts of IgE and IgG1 that are spontaneously found in the serum of unimmunized LAT Y175F+Y195F+Y235F mice.

20

Description

In this application, LAT Y136F, LAT Y175F, LAT Y195F, and LAT Y235F refer to the designated mutation itself, while LAT^{Y136F}, LAT^{Y175F}, LAT^{Y195F} and LAT^{Y235F} refer to mutants, mice
25 or products derived from these mutations.

Mutation of one or three tyrosine(s) among the four distal tyrosine of LAT protein (i.e. LAT Y136F, or LAT Y175F+Y195F+Y235F) is able to induce the development of
30 pathologies associated with exacerbated T_H2 immunity. Characteristics of the phenotype associated with this mutation are described in the following examples. Therefore, the present invention provides models of allergy and/or asthma or other diseases associated with T_H2 cell
35 deregulation or activity, more particularly T_H2 cell accumulation. Among the advantages of said models, it is found the rapidity of the model preparation (about 3-4 weeks

for a mice model instead of several months) and the exacerbated phenotype (for instance, exacerbated IgE production and tissue eosinophilia).

5 This phenotype due to the mutation of one or three tyrosine(s) among the four distal tyrosine residues of LAT protein (namely, LAT Y136F, or LAT Y175F+Y195F+Y235F) in mice was unpredictable, considering the phenotype of mice in which the four mutations are combined (LAT 4YF mice).
10 Indeed, LAT 4YF mice are totally devoid of thymocytes and T cells, because of the early differentiation blockage. Therefore, the LAT 4YF mice are unable to lead or suggest the phenotype observed for the LAT^{Y136F}, and LAT^{Y175F+Y195F+Y235F} mice. Moreover, none of the results of the previous studies
15 on cell lines suggests such a phenotype. Furthermore, the phenotype obtained in mice with the mutation Y136F could not be extrapolated in order to deduce the expected phenotype of mice having a composite mutation Y175F+Y195F+Y235F because of the different effects of the mutation Y136F and the
20 mutations Y175F, Y195F, and Y235F observed during the cell line studies.

The object of the present invention is to provide non-human animals having a mutated LAT gene of the invention leading
25 to an exaggerated T_H2 cell differentiation. By "gene" is intended cDNA or genomic sequence coding for a LAT protein. By "mutated LAT gene of the invention" is intended a LAT gene coding for a mutant LAT protein, the sequence of which corresponds to a wild type sequence and contains the
30 mutation of a single tyrosine among the four distal ones corresponding to Y136 in the mouse LAT protein or the composite mutation of the three distal tyrosine residues (corresponding to Y175, Y195 and Y235 in the mouse LAT protein). For example, the tyrosine corresponding to Y136 in
35 the mouse LAT protein is the residue Y132 in the human LAT protein. In a first preferred embodiment, said LAT gene coding for a mutant LAT protein contains a single mutation

of the tyrosine residue corresponding to Y136 in the mouse LAT protein. In a second preferred embodiment, said LAT gene coding for a mutant LAT protein contains the composite mutation of the three distal tyrosines, those corresponding 5 to Y175, Y195 and Y235 in the mouse LAT protein. Preferably, said non-human animals are mice and said non-human animals have the mutated gene coding for a mutant LAT protein, the sequence of which corresponds to a wild type sequence and contains the single mutation of the tyrosine residue at 10 position 136 or the composite mutation of the three distal tyrosine residues at positions 175, 195 and 235. Preferably, said mutation consists in the replacement by a residue preventing the association of the "tyrosine-based" sequences with the SH2 domain of proteins. More preferably, said 15 mutation consists in the replacement of the tyrosine by a phenylalanine (Y-F), an aspartic acid (Y-D) or a glutamic acid (Y-E). Still more preferably, said mutation consists in the replacement of the tyrosine by a phenylalanine (Y-F). Preferably, said non-human animals according to the 20 invention are mammals, and in particular, they are rodents. More preferably, said rodents are mice. Preferably, said animals are homozygous for the mutated LAT gene or are carrying a null allele of the LAT gene. Preferably, said mutated LAT gene is incorporated into the animal genome by 25 targeted insertion in order to keep said mutated LAT gene under the control of regulatory regions of the endogenous LAT gene.

By "distal" is intended the C-terminal end of the protein. 30 Therefore, the distal tyrosine residues are the tyrosines residues located at the C-terminal end of the protein.

In particular, the invention concerns any germ cell and somatic cell from said animals or any progeny thereof 35 containing the mutated LAT gene of the invention. More particularly, germ cells and somatic cells of said animals contain the mutated LAT gene of the invention as a result of

chromosomal incorporation into the animal genome, or into an ancestor of said animal. Preferably, said mutated LAT gene is incorporated into the animal genome by targeted insertion (homologous recombination) in order to keep said mutated LAT
5 gene under the control of regulatory regions of the endogenous LAT gene.

Therefore, a further object of the invention is to provide a mutated mouse gene coding for a mutant LAT protein, the
10 sequence of which corresponds to a wild type sequence and contains the single mutation of the tyrosine Y136, or a composite mutation of the tyrosine residues at positions 175, 195, and 235. Said mutation consists in the replacement by a residue preventing the association of the "tyrosine-
15 based" sequences with the SH2 domain of proteins. Preferably, said mutation of the tyrosine leads to its replacement by a phenylalanine, an aspartic acid or a glutamic acid. More preferably, said mutation of the tyrosine leads to its replacement by a phenylalanine. In a
20 preferred embodiment, the sequence of the gene encoding mutated mouse LAT^{Y136F} protein corresponds to sequence ID N°1. The invention further includes chromosomal DNAs containing exon 7 of the mutated gene (SEQ ID N°2). The invention concerns a mouse containing said mutated mouse gene.

25

The present invention also encompasses plasmids comprising a DNA or a part thereof, having a sequence corresponding to the mutated gene according to the invention. In a preferred embodiment, the plasmids of the invention contain a
30 restriction enzyme cleavage site, which is introduced in the intron 3' of exon 7. Advantageously, the restriction enzyme cleavage site is a Bgl II restriction site.

Said plasmids are useful for the generation of non-human
35 animals according to the present invention.

Consequently, the invention also encompasses non-human

embryos introduced with the plasmids of the invention, and non-human embryos obtained by homologous recombination using the plasmids of the invention. In a preferred embodiment, the non-human embryos are embryonic stem cells derived from a mouse. Advantageously, the ES cells are CK35 129/SV ES cells.

The invention also concerns the LAT mutant murine protein sequence containing the single mutation of the tyrosine Y136 or a composite mutation of the tyrosine residues at positions 175, 195, and 235. Said mutation consists in the replacement by a residue preventing the association of the "tyrosine-based" sequences with the SH2 domain of proteins. Preferably, said mutation of the tyrosine leads to its replacement by a phenylalanine, an aspartic acid or a glutamic acid. More preferably, said mutation of the tyrosine leads to its replacement by a phenylalanine. In one embodiment, the invention concerns a mutated LAT protein containing the mutated amino acid sequence of exon 7 (SEQ ID N°3).

The magnified and accelerated sequence of pathological events observed in the LAT ^{Y136F}, and LAT ^{Y175F+Y195F+Y235F} mice permits to readily start tests and studies. For example, mutant LAT ^{Y136F} mice phenotype is achieved when they are 4 weeks old.

The mutant non-human animal according to the invention are useful in various applications of interest, in particular:

- to analyze the impact of drugs on the molecular mechanisms that lead to exacerbated IgE production as well as tissue eosinophilia, and
- as a bioreactor allowing the dedicated production of IgE antibody of known specificity prior to or following a step of humanization of the mutated LAT mouse (preferably LAT ^{Y136F} or LAT ^{Y175F+Y195F+Y235F} mouse).

Consequently, the present invention provides models of allergy, and/or asthma disease comprising animals according to the invention. In particular, the animals of the invention can be used as models of eosinophilia and/or T_H2 cells deregulation, more particularly T_H2 cells accumulation.

Therefore, the invention concerns the use of a mutant non-human animal according to the present invention as a model of allergy and/or asthma disease. The invention also concerns the use of a mutant non-human animal according to the present invention as a model of eosinophilia. More generally, the invention concerns the use of a mutant non-human animal according to the present invention as a model of T_H2 cells deregulation, more particularly a model of T_H2 cells accumulation.

Due to the increased sensitivity of population, health difficulties such as asthma or allergies are more frequent. The animals according to the invention are suitable models to help the research in these domains.

Accordingly, the present invention provides a method of screening for a drug for treatment of allergy, asthma and/or disease associated with T_H2 cell deregulation or activity comprising the step of subjecting the animals according to the invention, which are administered with the drug to a comparison with said animals, not administered with the drug.

More particularly, the invention concerns a method of screening of drugs for treatment of allergy, asthma and/or disease associated with T_H2 cell deregulation or activity comprising the step of:

- 1) administering a candidate drug to a non-human animal having a LAT gene coding for a mutant LAT protein according to the present invention;
- 2) evaluating the effect of said drug on the symptom or sign

of allergy, asthma and/or disease associated with T_H2 cell deregulation or activity; and

3) selecting the drug that reduces said symptom or sign.

In a preferred embodiment, said screening method uses non-
5 human animals not administered with drugs as control experiments. In an other preferred embodiment, said effect of said drug can be evaluated by measuring at least one parameter selected from the group : IgE level, IgG1 level, interleukin level (preferably IL-4, IL-10, IL-5 and/or IL-
10 13), and eosinophilia. More preferably, said effect of said drug is evaluated by measuring the serum level of IgE and/or IgG1.

The invention also contemplates a method of screening drugs
15 for treatment of allergy, asthma and/or disease associated with T_H2 cell deregulation or activity comprising the step of:

1) subjecting cells having a LAT gene coding for a mutant LAT protein according to the present invention to a
20 candidate drug;

2) evaluating the effect of said drug on said cells;

3) selecting the drug having the desired effect.

In a preferred embodiment, said effect of said drug can be evaluated by measuring the interleukin production, more
25 particularly the IL-4 production.

An other object of this invention resides in a method of screening drugs that regulate the activity of T_H2 cells, comprising the step of:

30 1) administering a candidate drug to a non-human animal having a LAT gene coding for a mutant LAT protein according to the present invention; and

2) selecting a drug that modulates the activity of T_H2 cells in said non-human animal.

35

The screening methods can be used to select, identify, characterize and/or optimize candidate drugs. The candidate

drugs may be of any origin, nature and structure. Their concentration may be adjusted by the skilled artisan. Furthermore, several drugs may be tested in parallel, or in combination.

5

A further object of this invention is a method of producing a pharmaceutical composition for treating a disease associated with deregulated T_H2 cells activity, particularly asthma or allergy, the method comprising (i) selecting, 10 identifying, optimizing or characterizing a compound using a screening assay as described above and (ii) conditioning said compound, or a derivative thereof, in a pharmaceutically acceptable carrier or vehicle.

15 In still another application, the present invention provides bioreactors for a large-scale production of human IgE antibodies comprising the animals according to the invention. LAT^{Y136F} and LAT^{Y175F+Y195F+Y235F} mice are indeed able to produce tremendous amount of IgE, as it is described in 20 example 2. IgE produced by mutant mice are useful for applications such as desensitization or for kit of clinical assay.

Therefore, the invention concerns a method of production of 25 human IgE antibodies comprising the steps of :

- 1) providing a non-human animal expressing humanized IgE;
- 2) breeding said animal expressing humanized IgE with a non-human animal having a LAT gene coding for a mutant LAT protein according to the present invention;
- 30 3) immunizing the animal of the progeny with an allergen;
- 4) recovering humanized IgE specific to said allergen.

The step 4 can comprise the step of producing B cell hybridomas producing said humanized IgE specific to said allergen. The invention relates to said B cell hybridoma 35 producing said humanized IgE specific to said allergen.

Said non-human animal expressing humanized IgE can be

obtained by conventional knock-in in which the genetic segment corresponding to the constant exons of the IgE gene is substituted by the corresponding human sequence.

- 5 The invention concerns the non-human animal resulting from the breeding of the animal expressing humanized IgE with the non-human animal having a LAT gene coding for a mutant LAT protein according to the present invention.
- 10 The produced humanized IgE specific to said allergen can be used for desensitization and in clinical assays aiming at characterizing allergens, preferably atopic allergens, present in patient.
- 15 The invention contemplates the oligonucleotide probes specific to a mutated LAT gene coding for a mutant LAT protein containing the single mutation of the tyrosine corresponding to Y136 in the mouse LAT protein or a composite mutation of the three distal tyrosines
- 20 (corresponding to Y175, Y195 and Y235 in the mouse LAT protein). Such probes are useful to detect the presence of the mutation in a LAT gene. Hence, the invention provide oligonucleotides, the sequence of which corresponds to SEQ ID N°4 and/or SEQ ID N°5 as probes to screen the presence of
- 25 the mutation Y136 in the mouse LAT gene. More particularly, the invention concerns oligonucleotide probes specific to a mutated human LAT gene coding for a mutant LAT protein containing a single mutation of the tyrosine Y132 or a composite mutation of the tyrosine residues Y171, Y191 and
- 30 Y226. Such probes are useful for the detection of mutant LAT gene involved in asthma, allergy, eosinophilia and/or any disease associated with a T_H2 cells deregulation or activity. Said probes can be part of a diagnostic kit.
- 35 Therefore, the invention relates to a diagnostic method for asthma, allergy, eosinophilia and/or T_H2 cells deregulation, more particularly T_H2 cells accumulation, comprising the

detection of a mutated LAT gene coding for a mutant LAT protein containing a single mutation of Y132 or a composite mutation Y171+Y191+Y226. Additionally, the invention concerns a diagnostic kit for asthma, allergy, eosinophilia and/or T_H2 cells deregulation, more particularly T_H2 cells accumulation, comprising oligonucleotide probes for the detection of a mutated LAT gene coding for a mutant LAT protein containing a single mutation of Y132 or a composite mutation Y171+Y191+Y226.

10

Other characteristics and advantages of the invention are given in the following examples with reference to figures 1 to 11.

15 **EXAMPLES****Mutation LAT^{Y136}****Example 1: Production of mutant mice**

To test *in vivo* the importance of LAT^{Y136}, the inventors generated knock-in mice with a mutation replacing Y136 with phenylalanine (Y136F).

1. Materials and methods25 **Mice**

Mice were maintained in a specific pathogen-free animal facility.

LAT^{Y136F} mutation.

30 LAT genomic clones were isolated from a 129/Ola phage library. After establishing the nucleotide sequence and the exon-intron structure of the LAT gene, the tyrosine residue found at position 136 and encoded by exon 7 was mutated to phenylalanine. Mutagenesis was performed on a 1717-bp Eco RI-Xba I fragment encompassing part of exon 5, exons 6, 7
35 and 8. In addition to the intended mutation, a new Bgl II

restriction enzyme cleavage site was introduced in the intron 3' of exon 7 to accommodate the LoxP-flanked neo^r gene and facilitate subsequent identification of LAT^{Y136F} mutant mice. Finally, the targeting construct was extended to give 5 1.7 kb and 4.8 kb of homologous sequences 5' and 3' of the EcoRI-XbaI fragment, respectively (see Fig. 2). After electroporation of CK35 129/SV ES cells (C. Kress et al., 1998), and selection in G418, colonies were screened for homologous recombination by Southern blot analysis. The 5' 10 single-copy probe is a 0.9-kb Bgl II-Xba I fragment isolated from a LAT genomic clone. When tested on Bgl II-digested DNA, the 5' probe hybridizes either to a 8.5 kb wild-type fragment or to a 4.5 kb recombinant fragment. Homologous recombination events at the 3' side were screened by long 15 range PCR. Homologous recombinant ES clones were further checked for the presence of the intended mutation by sequencing the genomic segment corresponding to exon 7. Finally, a neo probe was used to ensure that adventitious non homologous recombination events had not occurred in the 20 selected clones.

Production of mutant mice.

Mutant ES cells were injected into Balb/c blastocysts. Two LAT^{Y136F} recombinant ES cell clones were found capable of germ 25 line transmission. The two mutant mouse lines were first bred to Deleter mice (Schwenk. F et al., 1995) to eliminate the Lox P-flanked neomycin cassette, and intercrossed to produce homozygous mutant mice. The two independently-derived mutant lines showed indistin-guishable phenotype. To 30 confirm that the LAT Y136F mutation had been genuinely introduced, LAT transcripts were cloned by reverse transcription and PCR amplification from the thymus of the mutated mice, and the presence of the intended mutation confirmed by DNA sequence analysis. Screening of mice for 35 the presence of the LAT Y136F mutation was performed by PCR using the following pairs of oligonucleotides:

c : 5'-GTGGCAAGCTACGAGAACCAGGGT-3' (SEQ ID N°4);

d : 5'-GACGAAGGAGCAAAGGTGGAAGGA-3' (SEQ ID N°5).

The single Lox P site remaining in the LAT Y136F allele after deletion of the neo^F resulted in an amplified PCR product 140 bp-longer than the 510 bp-long fragment amplified from the wild-type LAT allele.

2) Mutant mice development

Mice homozygous for the LAT^{Y136F} mutation, hereafter denoted LAT^{Y136F}, were born at expected Mendelian frequencies and their T cells contained levels of LAT proteins similar to wild-type T cells. At birth LAT^{Y136F} mice displayed peripheral lymphoid organs of normal size. Beginning at about 3 weeks, however, the spleen and lymph nodes of the mutant mice started to enlarge relative to wild-type littermates, such that by 15 weeks of age, spleen cellularity was approximately 10 times that of wild-type mice (Fig.3 A-C). Despite marked lymphocytic infiltrations in the lung, liver and kidney, homozygotes lived to at least 17 weeks of age, and no chronic intestinal inflammation or tumor formation was observed. The effects of the LAT^{Y136F} mutation were only detectable after breeding mice to homozygosity or to mice carrying a null allele of the LAT gene.

Example 2: Effect of the mutation: spontaneous exaggerated T helper type 2 immunity in mice

1. Materials and methods

Purification of CD4⁺ T cells and eosinophils.

Lymph node and spleen cells from several mice were pooled and the CD4⁺ cells purified using a high gradient magnetic cell separation system (S. Miltenyi et al., 1990). Eosinophils were sorted on a FACSvantageTM on the basis of their FSChigh, HSA+, and CD11b+ phenotype.

Antibodies and flow cytometric analysis.

Before staining, cells were preincubated on ice for at least 10 min with polyclonal mouse and rat Ig to block Fc receptors. Flow cytometric analysis was performed as 5 described previously (M. Malissen *et al.*, 1995). All the antibodies were from BD PharMingen except the anti-CCR3 antibody that was purchased from R&D.

Staining for intracellular cytokines.

10 Before intracellular cytokine staining, cells (1.5×10^6) were cultured for 4 h in the presence of monensin (GolgiStop; BD PharMingen) at a final concentration of 2 μ M. Cells were then immediately placed on ice, washed, resuspended in PBS 1X, 1% FCS, 0.20% sodium azide, and 15 stained with an APC-conjugated anti-CD4 antibody. For intracellular cytokine staining, cells were first fixed using the cytofix/cytoperm kit (BD PharMingen). Each cell sample was subsequently split into aliquots that were separately stained with (1) a combination of FITC-conjugated 20 anti-IFN- α and PE-conjugated anti-IL-2 antibodies, (2) a combination of FITC-conjugated anti-IL-5 and PE-conjugated anti-IL-4 antibodies, and (3) a combination of fluorochrome-conjugated and isotype-matched negative control Ig (BD PharMingen). After a final wash, CD4⁺ cells (10^4) were 25 analyzed on a FACSCalibur™ flow cytometer after gating out dead cells using forward and side scatters.

RNase protection assay.

For multiplex cytokine transcript analysis, total cellular 30 RNA was isolated from the specified cells using TRIzol (GIBCO-BRL Life Technologies) and analyzed by ribonuclease protection assay using an MCK-1 RiboQuant™ custom mouse template set (BD Pharmingen). Briefly, ³²P-labeled riboprobes were mixed with 10 μ g of RNA, incubated at 56°C for 12 to 16 35 hours, and then treated with a mixture of RNases A and T1 and proteinase K. RNase-protected ³²P-labeled RNA fragments

were separated on denaturing polyacrylamide gels and the intensity of the bands evaluated with a Fuji imaging plate system.

5 Determination of serum isotype-specific immunoglobulin levels.

The titres of polyclonal IgM, IgG1, IgG2a, IgG2b, IgG3 and IgA antibodies and κ and λ light chains were determined using isotype-specific ELISA (Southern Biotechnology). The
10 concentrations of IgG1 and IgE were determined by comparing test sample dilution series values with isotype control standards.

2. Results

15 A prominent phenotype of the CD4 T cells found in LAT^{Y136F} mice was revealed when the inventors measured their ability to make cytokines. Due to the short half-lives of cytokines and of their transcripts, their analysis generally requires restimulation of T cells in vitro with PMA and ionomycin. A
20 multiprobe RNase protection assay detecting levels of transcripts of 9 cytokines showed that CD4 T cells freshly isolated from LAT^{Y136F} mice contained sufficient IL-4 and IL-10 transcripts to allow their detection even without ex vivo restimulation (Fig. 4A). Upon activation by PMA/ionomycin
25 the levels of IL-4 and IL-10 transcripts they contained were further increased, and IL-5, IL-13, and IFN- α transcripts became readily detectable (Fig. 4B). In marked contrast, wild-type CD4 T cells yielded only the IL-2 and IFN- α transcripts expected for primary T cells. Analysis of IL-4
30 production at the single cell level, showed that following a 4 hr activation with PMA/ionomycin, close to 80% of the CD4 T cells isolated from LAT^{Y136F} mice expressed very high levels of IL-4 (Fig. 4C). Consistent with the notion that these CD4 T cells were refractory to TCR stimuli, none of them scored
35 as IL-4+ in response to TCR cross-linking (Fig. 4C). Thus, LAT Y136F spontaneously developed a high frequency of T_H2

cells. In the case of wild-type CD4 T cells, T_H2 polarization of such magnitude is only achieved following prolonged antigenic stimulation in the presence of IL-4.

5 Light scatter analysis of thymic and lymph node cells from LAT^{Y136F} mice older than 4 weeks revealed a unique cell population that was almost absent from age-matched wild-type mice, and showed both an intermediate forward scatter and a high side scatter (Fig. 5A, 5B, 6A). Based on several of
10 criteria, these cells were identified as eosinophils (Fig. 6). Minute numbers of eosinophils normally reside in wild-type thymi, and their augmentation in LAT^{Y136F} thymi may primarily result from an intrinsic expression of LAT^{Y136F} molecules. However, LAT transcripts were undetectable in
15 eosinophils purified from LAT^{Y136F} mice, meaning that the thymic and lymph node eosinophilia they manifest result from the production of IL-5 by the abnormal CD4 cells present in these mutant mice.

20 Secondary lymphoid organs of 6-week old LAT^{Y136F} mice contained 7 to 10 times more B cells than their wild-type counterparts. Thus, the splenomegaly and generalized lymphadenopathy that developed in young LAT^{Y136F} mice can be mostly accounted for by cells belonging to the T and B cell
25 lineages. Over 90% of the mature B cells found in the spleen and lymph nodes of 6-week old wild type littermates had a resting phenotype (Fig. 7A). In marked contrast, only 25% of the B cells found in the enlarged secondary lymphoid organs of age-matched LAT^{Y136F} littermates showed a resting
30 phenotype. Among the remaining B cells, 25% showed an hyperactivated phenotype, and 50% expressed a phenotype typical of antibody producing cells. Coincident with the presence of these latter cells, serum IgG1 concentrations were elevated approximately 200 times compared to wild-type
35 mice, whereas those of IgE were elevated 2500 to 10000 times (Fig. 7C). In contrast, the levels of the other Ig isotypes did not differ significantly from those of wild-type serum

(Fig. 7B). In support of a polyclonal hypergammaglobulinemia G1 and E, the concentrations of kappa and lambda light chains were both markedly augmented in the serum of LAT^{Y136F} mice (Fig. 7B). Notably, IgE and IgG1 antibody concentrations reached a plateau as early as 5 weeks of age (Fig. 7C), the values of which exceeded the extraordinarily large amounts of IgE and IgG1 previously reported for mice deprived of NFATc2 and NFATc3 transcription factors. Given that B cells do not express LAT proteins, and considering that isotype switching to IgE and IgG1 is highly dependent on the presence of IL-4 and IL-13, the overproduction of IgE and IgG1 noted in LAT^{Y136F} mice is secondary to the presence of an abnormally high frequency of T_H2 effectors.

15 **Example 3: Production of IgE**

Mice expressing humanized IgE are developed by conventional knock-in strategy in which the genetic segment corresponding to the constant exons of the IgE gene is substituted by the corresponding human sequence. Mice with a humanized IgE locus are bred into LAT Y136F mice. Following immunization, B cell hybridomas producing specific human IgE are produced, and the resulting specific human IgE are used as "standard" in clinical assays aiming at characterizing atopic allergens present in patients."

25

Example 4: Screening for a drug

Mutant mice and control ones will be treated with a variety of drugs or original compounds. Their effects will be analyzed in vivo by measuring various parameters such as:

- 30
- T_H2 cells differentiation.
 - Production of T_H2 types cytokines
 - Eosinophilia
 - Hypergammaglobulinemia G1 and E .

Example 5: Production of mutant mice

To test *in vivo* the importance of the three carboxy-terminal tyrosines (LAT Y175, LAT Y195 and LAT Y235), the inventors generated knock-in mice with a mutation replacing these three tyrosines with phenylalanine (LAT Y175F + Y195F + Y235F).

1. Materials and methods**Mice**

Mice were maintained in a specific pathogen-free animal facility.

LAT Y175F + Y195F + Y235F mutation.

LAT genomic clones were isolated from a 129/Ola phage library. After establishing the nucleotide sequence and the exon-intron structure of the LAT gene (EMBL Nucleotide Sequence Database; accession number: AJ438435), the tyrosine residues found at positions 175, 195 and 235 and encoded by exons 9, 10, 11 were mutated to phenylalanine. Mutagenesis was performed on a 815 bp NcoI-BamHI fragment encompassing exons 9, 10, 11 (coding for tyrosines 175 (exon 9), 195 (exon 10) and 235 (exon 11) and part of exon 12 (corresponding to the 3' untranslated region of LAT). Each exon was mutated independently and new restriction sites were introduced for facilitating subsequent cloning steps. A new Eco RI site was introduced on the 5' side of the NcoI site, BamHI and ClaI sites were introduced between exons 9 and 10, a HindIII site was introduced between exons 10 and 11, and BglII, XhoI, and NotI sites were introduced on the 3' side of exon 11 in lieu of the original BamHI site. PCR reactions were performed with Pwo DNA polymerase (Boehringer Mannheim), and PCR products were purified and cut with with EcoRI and BamHI for exon 9, BamHI and HindIII for exon 10,

and HindIII and NotI for exon 11. These three fragments were assembled in a pBS-KS plasmid (Stratagene). The resulting plasmid was used to clone a 3.5 kb Eco RI-Nco I genomic fragment providing a 5' homology arm and a 4.3 kb-Sal I genomic fragment providing a 3' homology arm. Finally a loxP flanked neo^r gene was introduced using the BamHI and ClaI sites that were engineered between exons 9 and 10. After electroporation of CK35 129/SV embryonic stem (ES) cells and selection in G418, colonies were screened for homologous recombination by southern blot analysis using a 3' single-copy probe that consisted of a 1.1 kb EcoRI-HindIII fragment isolated from a LAT genomic clone. When tested on BamHI digested genomic DNA, the 3' probe hybridizes either to a 7.0 kb wild-type fragment or to a 9.1 kb recombinant fragment. The presence of a genuine recombination event was checked by PCR using the following pair of primers (depicted in Figure 8):

f: 5'-CCCAGAGGCAAACCTCTGAAG-3' (SEQ ID N°6) and

g: 5'-TCGAATTCGCCAATGACAAGACGC-3' (SEQ ID N°7). This PCR gives a band of 8.6 kb in the recombinant ES clones only. Homologous recombinant ES clones were further checked for the presence of the intended mutations by sequencing the genomic segment corresponding to exons 9, 10 and 11. Finally, a neo probe was used to ensure that adventitious non-homologous recombination events had not occurred in the selected clones.

Production of mutant mice.

Mutant ES cells were injected into Balb/c blastocysts. Two LAT Y175F + Y195F + Y235F recombinant ES cell clones were found capable of germ line transmission. The two mutant mouse lines were first bred to Deleter mice (Schwenk. F et al., 1995) to eliminate the Lox P-flanked neomycin cassette, and intercrossed to produce homozygous mutant mice. The two independently-derived mutant lines showed indistinguishable phenotype. To confirm that the LAT Y175F + Y195F + Y235F mutation had been genuinely introduced, LAT transcripts were

cloned by reverse transcription and PCR amplification from the thymus of the mutated mice, and the presence of the intended mutation confirmed by DNA sequence analysis. Screening of mice for the presence of the LAT Y175F + Y195F + Y235F mutation was performed by PCR using the following pairs of oligonucleotides:

d: 5'-GGAGACTTAGATGTCTGAGCCG-3' (SEQ ID N°8) and

e: 5'-GACAGACCAGCAGGGACAGTG-3' (SEQ ID N°9) (Wt 238bp, mutant 435bp).

10 The single Lox P site remaining in the LAT Y175F + Y195F + Y235F allele after deletion of the neo^r resulted in an amplified PCR product 140 bp-longer than the 510 bp-long fragment amplified from the wild-type LAT allele.

15 2) Mutant mice development

Mice homozygous for the LAT Y175F + Y195F + Y235F mutation, hereafter denoted LAT Y175F + Y195F + Y235F were born at expected Mendelian frequencies and their T cells contained levels of LAT proteins similar to wild-type T cells. At birth LAT Y175F + Y195F + Y235F mice displayed peripheral lymphoid organs of normal size. Beginning at about 3 months, however, the spleen and lymph nodes of the mutant mice started to enlarge relative to wild-type littermates, such that by 3 months of age, spleen cellularity was approximately 5 times that of wild-type mice. Homozygotes lived to at least 5 months of age, and no chronic intestinal inflammation or tumor formation was observed. The effects of the LAT Y175F + Y195F + Y235F mutation were only detectable after breeding mice to homozygosity or to mice carrying a null allele of the LAT gene.

Example 6: Effect of the mutation: a subset of gamma/delta T cells expands and acquire a spontaneous exaggerated T helper type 2 immunity in mice.

35

1. Materials and methods

Purification of gamma/delta T cells and eosinophils.

Spleen cells from several mice were pooled and the gamma/delta T cells purified using a high gradient magnetic cell separation system (S. Miltenyi et al., 1990).

Antibodies and flow cytometric analysis.

Before staining, cells were preincubated on ice for at least 10 min with polyclonal mouse and rat Ig to block Fc receptors. Flow cytometric analysis was performed as described previously (M. Malissen et al., 1995). All the antibodies were from BD PharMingen.

Staining for intracellular cytokines.

Before intracellular cytokine staining, cells (1.5×10^6) were cultured for 4 h in the presence of monensin (GolgiStop; BD PharMingen) at a final concentration of 2 μ M. Cells were then immediately placed on ice, washed, resuspended in PBS 1X, 1% FCS, 0.20% sodium azide, and stained with an APC-conjugated anti-CD5 antibody. For intracellular cytokine staining, cells were first fixed using the cytofix/cytoperm kit (BD PharMingen). Each cell sample was subsequently split into aliquots that were separately stained with (1) a combination of FITC-conjugated anti-IFN- γ and PE-conjugated anti-IL-2 antibodies, (2) a combination of FITC-conjugated anti-IL-5 and PE-conjugated anti-IL-4 antibodies, and (3) a combination of fluorochrome-conjugated and isotype-matched negative control Ig (BD PharMingen). After a final wash, CD5⁺ cells (10^4) were analyzed on a FACSCaliburTM flow cytometer after gating out dead cells using forward and side scatters.

RNase protection assay.

For multiplex cytokine transcript analysis, total cellular RNA was isolated from the specified cells using TRIzol (GIBCO-BRL Life Technologies) and analyzed by ribonuclease protection assay using an MCK-1 RiboQuantTM custom mouse

template set (BD Pharmingen). Briefly, ^{32}P -labeled riboprobes were mixed with 10 μg of RNA, incubated at 56°C for 12 to 16 hours, and then treated with a mixture of RNases A and T1 and proteinase K. RNase-protected ^{32}P -labeled RNA fragments were separated on denaturing polyacrylamide gels and the intensity of the bands evaluated with a Fuji imaging plate system.

Determination of serum isotype-specific immunoglobulin levels.

The titres of polyclonal IgM, IgG1, IgG2a, IgG2b, IgG3 and IgA antibodies and κ and λ light chains were determined using isotype-specific ELISA (Southern Biotechnology). The concentrations of IgG1 and IgE were determined by comparing test sample dilution series values with isotype control standards.

2. Results

A prominent phenotype of the CD90.2^+ , CD5^+ gamma/delta T cells found in LAT Y175F + Y195F + Y235F mice (see Figure 9) was revealed when the inventors measured their ability to make cytokines. Due to the short half-lives of cytokines and of their transcripts, their analysis generally requires restimulation of T cells in vitro with PMA and ionomycin. A multiprobe RNase protection assay detecting levels of transcripts of 9 cytokines showed that gamma/delta T cells freshly isolated from LAT Y175F + Y195F + Y235F mice contained large amounts of IL-4, IL-5, IL-10 and IL-13 transcripts to (Figure 10). This attribute is reminiscent of the observation made with the alpha/beta T cells present in the periphery of the LAT Y136F mice. In marked contrast, wild-type CD4^+ T cells yielded only the IL-2 and IFN- γ transcripts expected for primary T cells. Analysis of IL-4 production at the single cell level, showed that following a 4 hr activation with PMA/ionomycin, close to 80% of the CD4^+ T cells isolated from LAT Y175F + Y195F + Y235F mice expressed very high levels of IL-4. Thus, LAT Y175F + Y195F

+ Y235F mice spontaneously developed a high frequency of gamma/delta T cells with a Th2 phenotype. In the case of wild-type CD4 T cells, Th2 polarization of such magnitude is only achieved following prolonged antigenic stimulation in the presence of IL-4.

The spleen of 3-month old LAT Y175F + Y195F + Y235F mice contained 5 to 10 times more B cells than their wild-type counterparts. Thus, the splenomegaly that developed in LAT Y175F + Y195F + Y235F mice can be mostly accounted for by cells belonging to the T and B cell lineages. Over 90% of the mature B cells found in the spleen and lymph nodes of 3-month old wild type littermates had a resting phenotype (Fig. 11A). In marked contrast, only 16% of the B cells found in the enlarged secondary lymphoid organs of age-matched LAT Y175F + Y195F + Y235F littermates showed a resting phenotype. Among the remaining B cells, 21% showed an hyperactivated phenotype, and 63% expressed a phenotype typical of antibody producing cells. Coincident with the presence of these latter cells, serum IgG1 concentrations were elevated approximately 100 times compared to wild-type mice, whereas those of IgE were elevated 500 to 5000 times (Fig. 11). In contrast, the levels of the other Ig isotypes did not differ significantly from those of wild-type serum. Given that mature B cells do not express LAT proteins, and considering that isotype switching to IgE and IgG1 is highly dependent on the presence of IL-4 and IL-13, the overproduction of IgE and IgG1 noted in LAT Y175F + Y195F + Y235F mice is secondary to the presence of an abnormally high frequency of gamma/delta T cells producing Th2 cytokines.

Example 7: Production of IgE

Mice expressing humanized IgE are developed by conventional knock-in strategy in which the genetic segment corresponding to the constant exons of the IgE gene is substituted by the corresponding human sequence. Mice with a humanized IgE

locus are bred into LAT Y175F + Y195F + Y235F mice. Following immunization, B cell hybridomas producing specific human IgE are produced, and the resulting specific human IgE are used as "standard" in clinical assays aiming at
5 characterizing atopic allergens present in patients."

Example 8: Screening for a drug

Mutant mice and control ones will be treated with a variety of drugs or original compounds. Their effects will be
10 analyzed in vivo by measuring various parameters such as:

- T_H2 cells differentiation.
- Production of T_H2 types cytokines
- Hypergammaglobulinemia G1 and E .

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Claims

1. A non-human animal having a mutated LAT gene coding for a mutant LAT protein, wherein said mutant LAT protein leads to an exaggerated T_H2 cell differentiation.
- 5 2. The non-human animal according to claim 1, wherein the sequence of said mutant LAT protein corresponds to a wild type sequence and contains a single mutation of the tyrosine corresponding to Y136 in the mouse LAT protein.
- 10 3. The non-human animal according to claim 2, wherein said mutated LAT gene coding for a mutant LAT protein comprises exon 7 of the mutated gene (SEQ ID No 2).
- 15 4. The non-human animal according to claim 1, wherein the sequence of said mutant LAT protein contains a composite mutation of the three distal tyrosine residues.
5. The non-human animal according to any of claims 1 to 4,
20 wherein said non-human animal is a mammal.
6. The non-human animal according to claim 5, wherein said mammal is a rodent.
- 25 7. The non-human animal according to claim 6, wherein said rodent is a mouse.
8. The non-human animal according to any of claims 1 to 7, wherein said mutation consists in the replacement of the
30 tyrosine by a residue preventing the association of the "tyrosine-based" sequences with the SH2 domain of proteins.
9. The non-human animal according to any of claims 1 to 7, wherein said single mutation consists in the replacement of
35 the tyrosine by a phenylalanine (Y-F), an aspartic acid (Y-

D) or a glutamic acid (Y-E).

10. The non-human animal according to claim 9, wherein said single mutation consists in the replacement of the tyrosine
5 by a phenylalanine (Y-F).

11. The non-human animal according to any of claims 1 to 10, wherein said non-human animal is homozygous for the mutated LAT gene or carries a null allele of the LAT gene.

10

12. The non-human animal according to any of claims 1 to 11, wherein said mutated LAT gene is incorporated into the animal genome by targeted insertion in order to keep said mutated LAT gene under the control of regulatory regions of
15 the endogeneous LAT gene.

13. A germ cell or somatic cell from a non-human animal according to any one of claims 1-12 or any progeny thereof containing the mutated LAT gene.

20

14. Use of an animal according to any of claims 1 to 12 as a model of allergy, asthma, eosinophilia and/or disease associated with T_H2 cell deregulation.

25 15. A method of screening for a drug for treatment of allergy, asthma and/or disease associated with T_H2 cell deregulation comprising the step of subjecting animals according to any of claims 1 to 12 which are administered with the drug to a comparison with said animals, not
30 administered with the drug.

16. A method of screening for drugs for treatment of allergy, asthma and/or disease associated with T_H2 cell deregulation comprising the step of:

- 35 1) administering a candidate drug to a non-human animal according to any of claims 1 to 12;
2) evaluating the effect of said drug on the symptom or sign

of allergy, asthma and/or disease associated with T_H2 cell deregulation; and

3) selecting the drug that reduces said symptom or sign.

5 17. The method according to claim 16, wherein said effect of said drug can be evaluated by measuring at least one parameter selected from the group of IgE level, IgG1 level, interleukin level (IL-4, IL-10, IL-5 and/or IL-13), and eosinophilia.

10

18. The method according to claim 17, wherein said effect of said drug can be evaluated by measuring the serum level of IgE and/or IgG1.

15 19. A method of screening drugs for treatment of allergy, asthma and/or disease associated with T_H2 cell deregulation comprising the step of:

1) subjecting cells according to claim 13 to a candidate drug;

20 2) evaluating the effect of said drug on said cells;

3) selecting the drug having the desired effect.

20. A method of screening for drugs for that regulate the activity of T_H2 cells, comprising the step of:

25 1) administering a candidate drug to a non-human animal according to any of claims 1 to 12; and

2) selecting a drug that modulates the activity of T_H2 cells in said non-human animal.

30 21. A method of producing a pharmaceutical composition for treating a disease associated with deregulated T_H2 cells activity, particularly asthma or allergy, the method comprising (i) selecting, identifying, optimizing or characterizing a compound using a screening method according
35 to any of claims 16 to 20 and (ii) conditioning said compound, or a derivative thereof, in a pharmaceutically acceptable carrier or vehicle.

22. A bioreactor for a large scale production of human IgE antibodies comprising an animal according to any of claims 1 to 12.

5

23. A method of production of human IgE antibodies comprising the steps of :

- 1) providing a non-human animal expressing humanized IgE;
- 2) breeding said animal expressing humanized IgE with a non-
10 human animal according to any of claims 1 to 12;
- 3) immunizing the animal of the progeny with an allergen;
- 4) recovering humanized IgE specific to said allergen.

24. The method according to claim 23, wherein the step 4
15 comprises the step of producing B cell hybridomas producing said humanized IgE specific to said allergen.

25. A B cell hybridoma obtained by the method according to claim 24.

20

26. A mutated mouse gene coding for a mutant LAT protein, the sequence of which corresponds to a wild type sequence and contains a single mutation of the tyrosine Y136.

25 27. A mutated mouse gene coding for a mutant LAT protein, the sequence of which corresponds to a wild type sequence and contains a composite mutation of the three distal tyrosine residues.

30 28. The mouse gene according to any of claims 26 to 27, wherein said mutation consists in the replacement of the tyrosine by a phenylalanine (Y-F), an aspartic acid (Y-D) or a glutamic acid (Y-E).

35 29. The mouse gene according to 28, wherein said mutation consists in the replacement of the tyrosine by a phenylalanine (Y-F).

30. The mouse gene according to 26, wherein the sequence corresponds to sequence ID N°1.

5 31. The mouse gene according to 30, wherein the sequence contains exon 7 of the mutated gene (SEQ ID N°2).

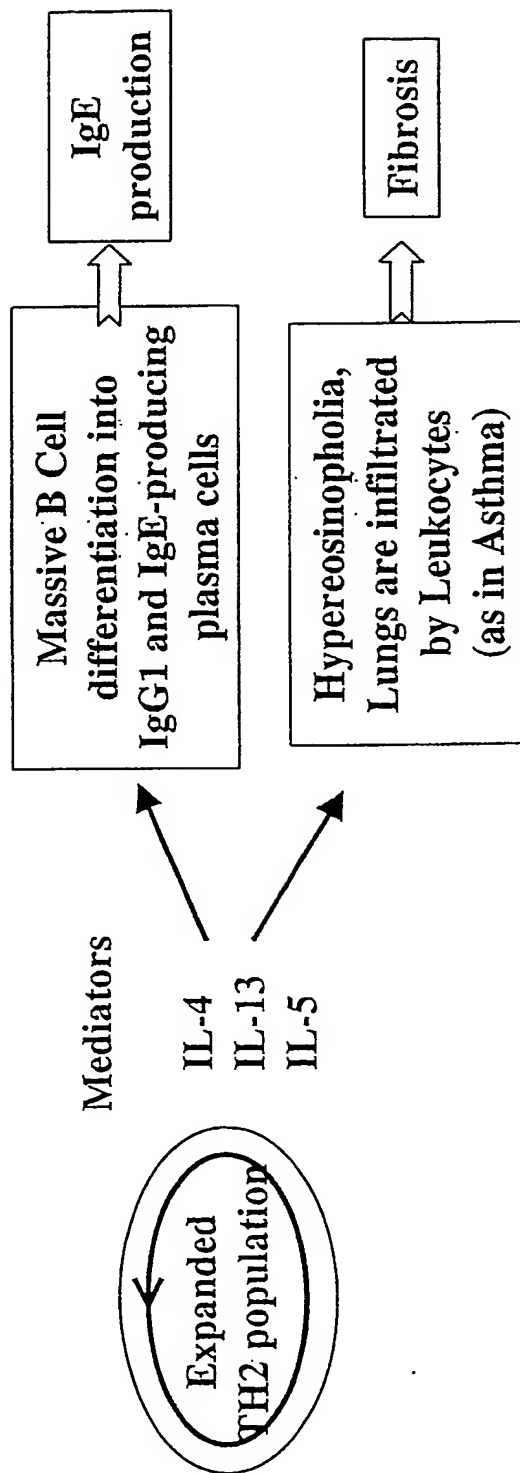
32. A diagnostic method for asthma, allergy, eosinophilia and/or T_H2 cells deregulation comprising the detection of a
10 mutated LAT gene coding for a mutant LAT protein containing a mutant LAT protein containing a single mutation of the tyrosine Y132 or a composite mutation of the three distal tyrosines Y171, Y191 and Y226.

15 33. A diagnostic kit for asthma, allergy, eosinophilia and/or T_H2 cells deregulation comprising oligonucleotide probes for the detection of a mutated LAT gene coding for a mutant LAT protein containing a single mutation of the tyrosine Y132 or a composite mutation of the three distal
20 tyrosines Y171, Y191 and Y226.

34. A non-human animal resulting from the breeding of a non-human animal expressing humanized IgE with the non-human animal according to any of claims 1 to 12.

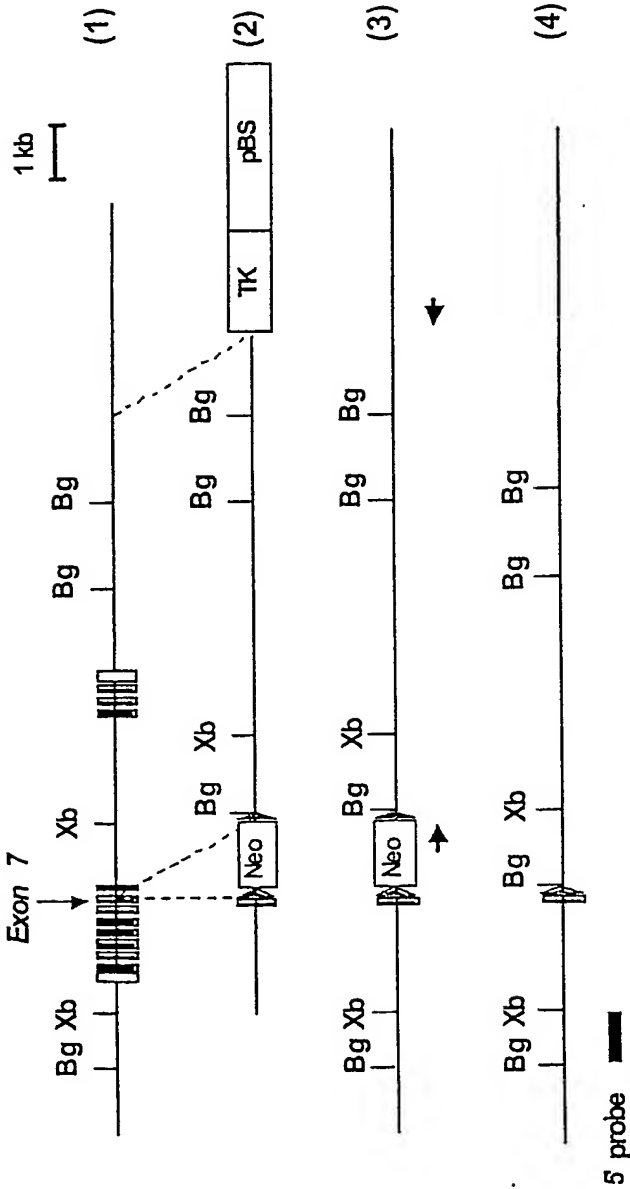
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Figure 1



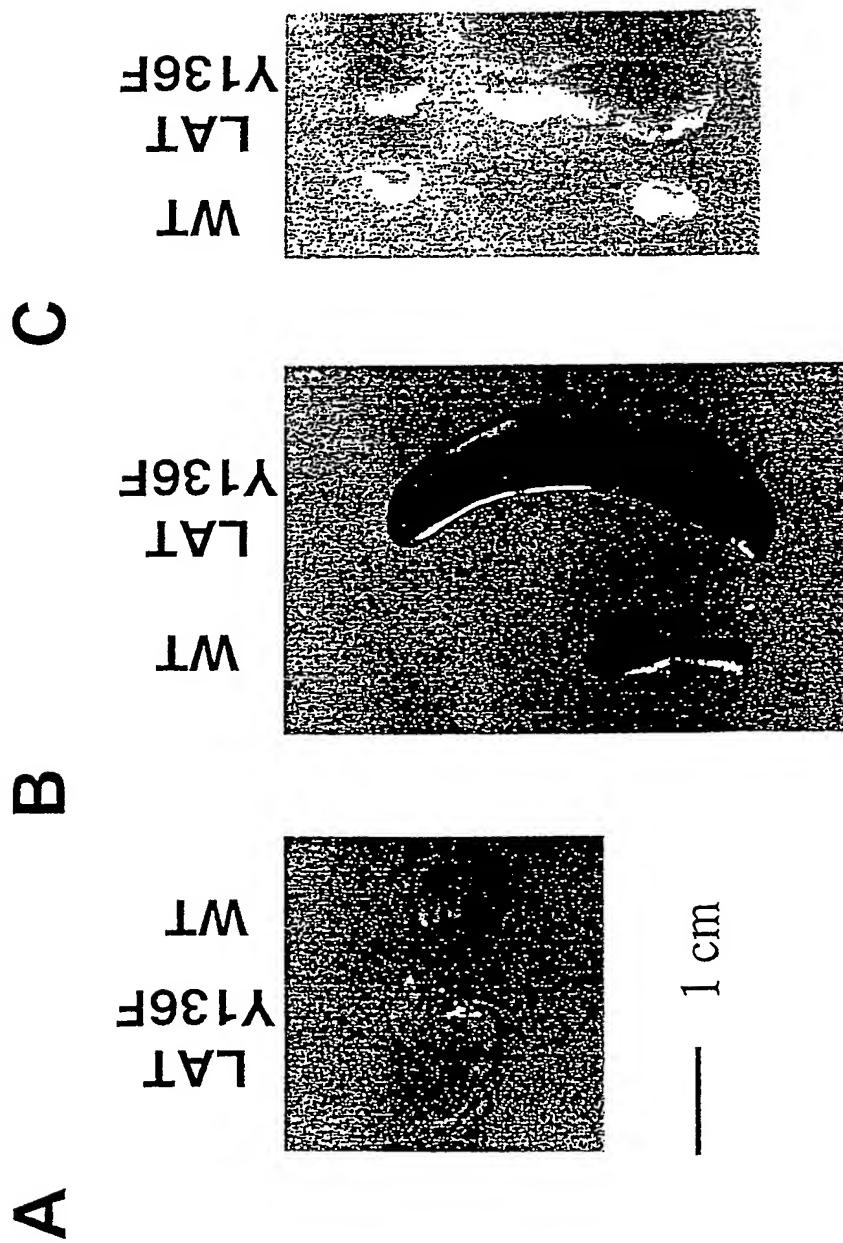
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Figure 2



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Figure 3



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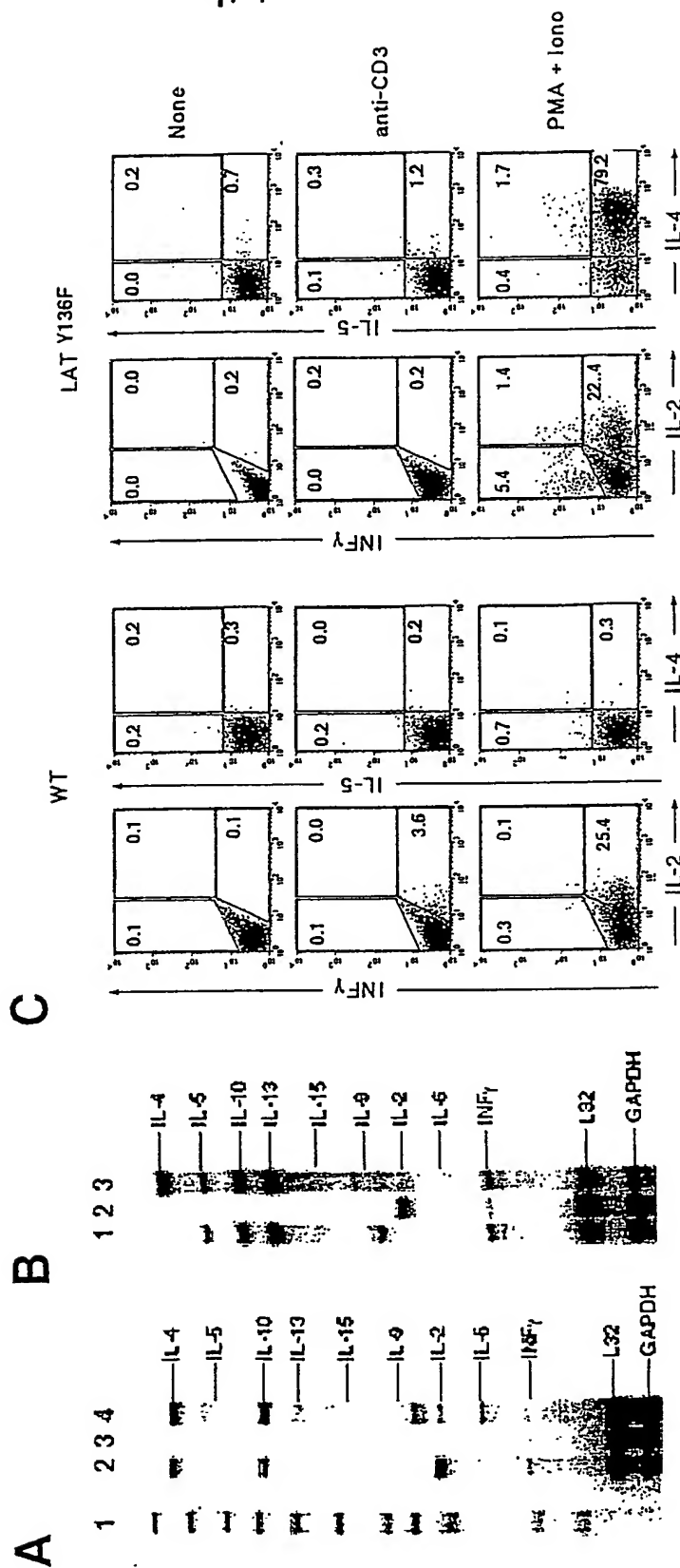
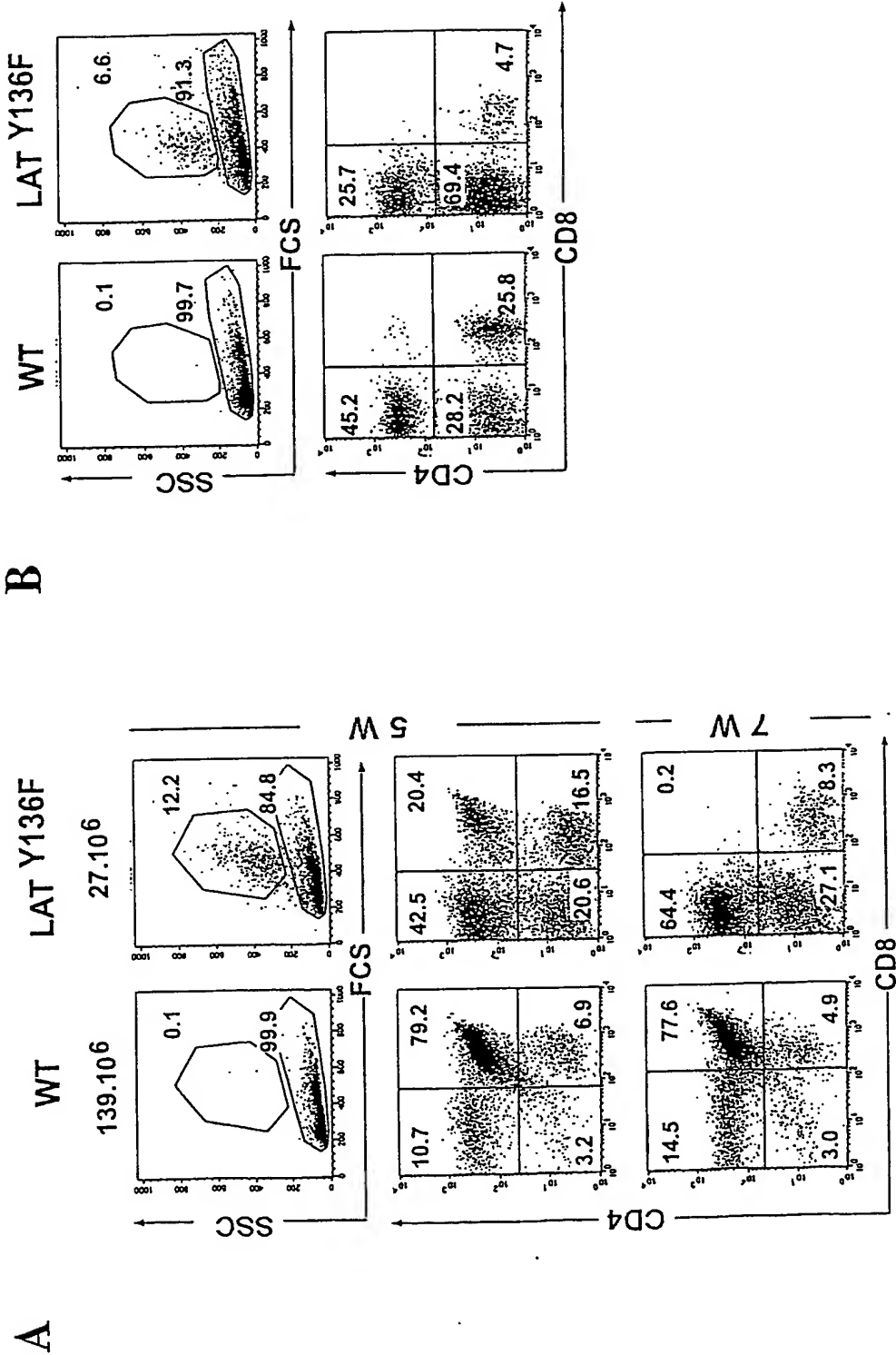
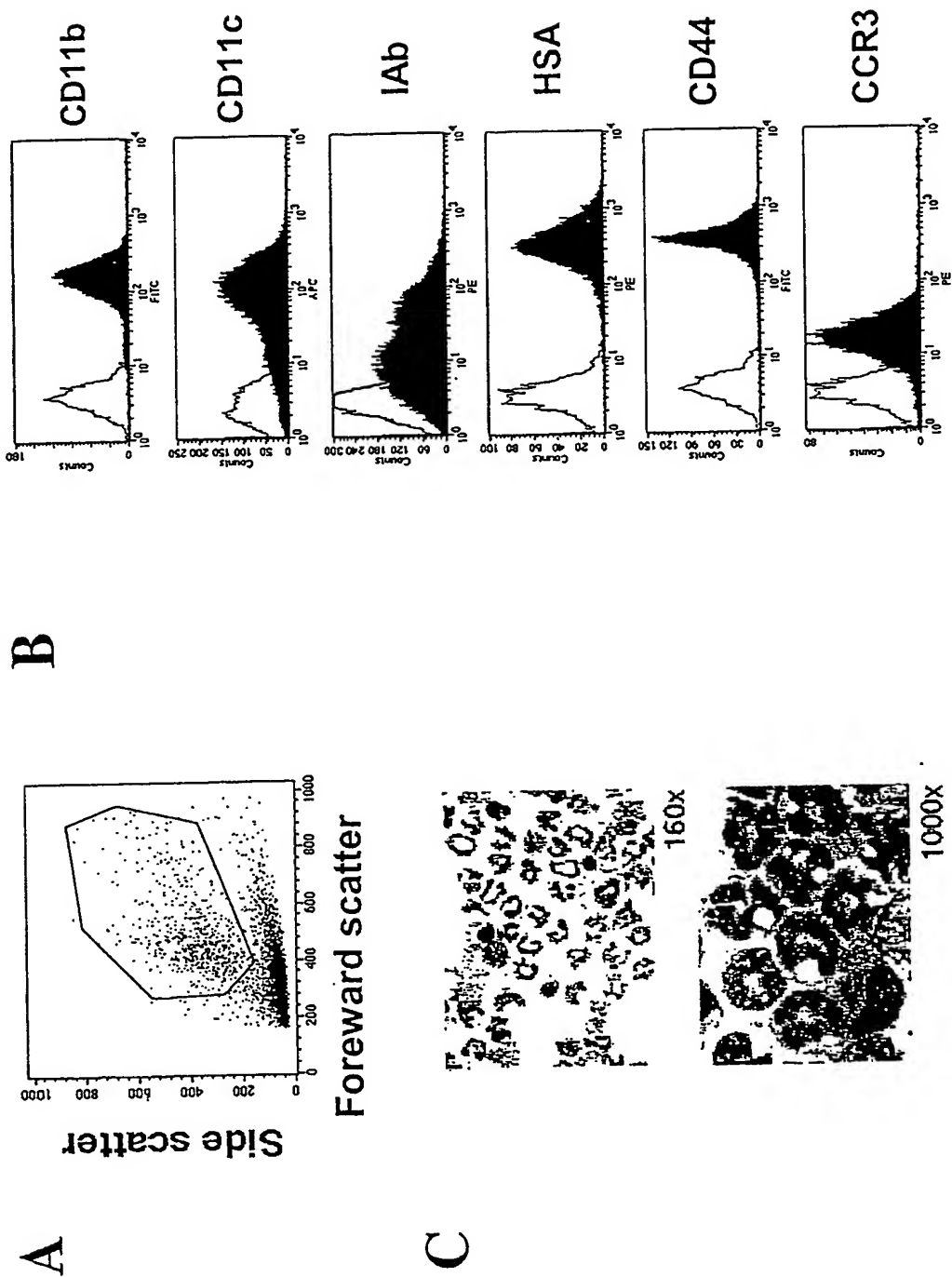


Figure 5



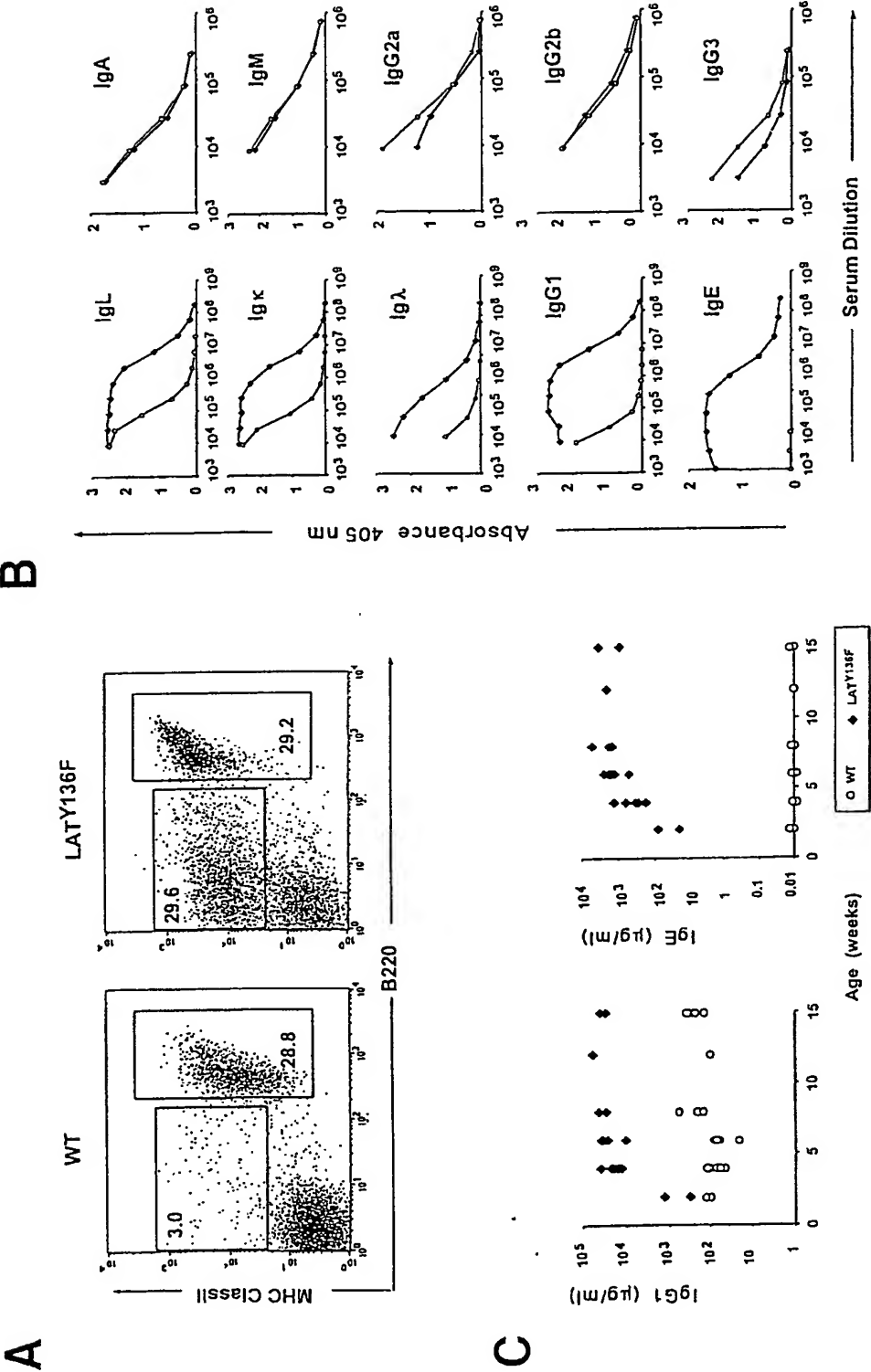
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Figure 6



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Figure 7



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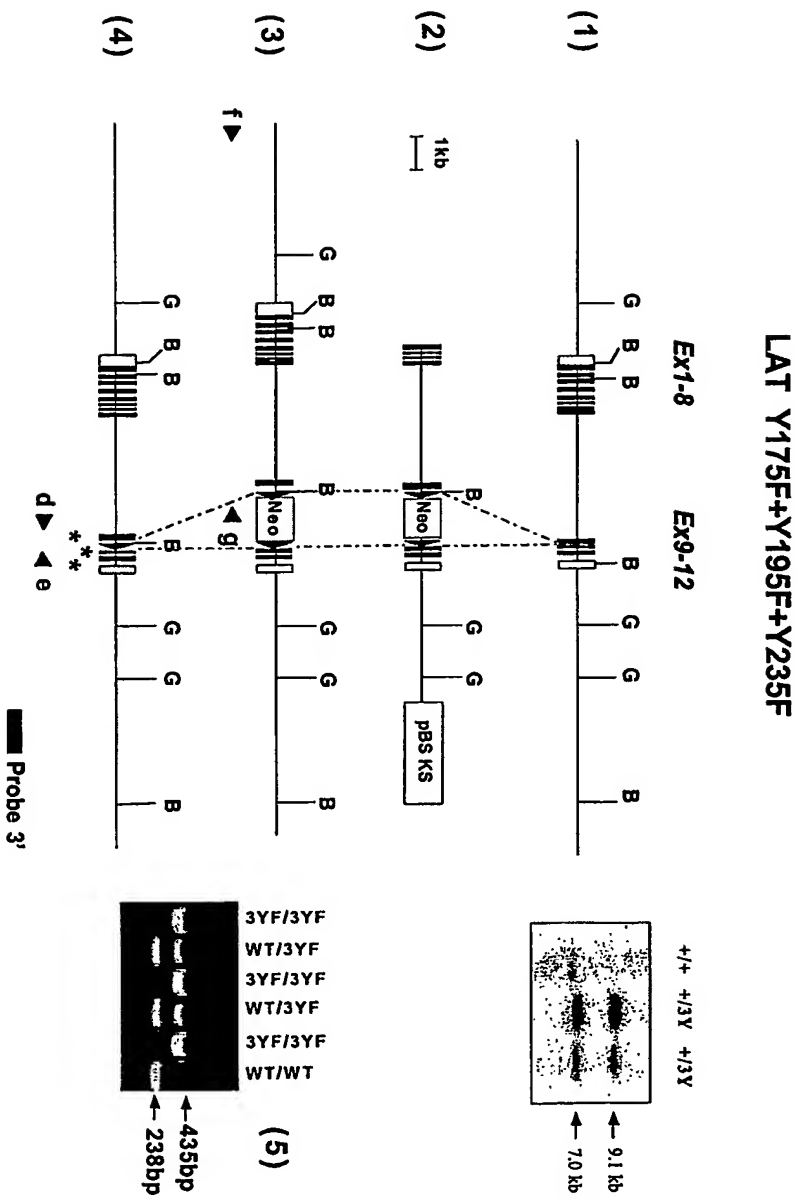


FIGURE 8

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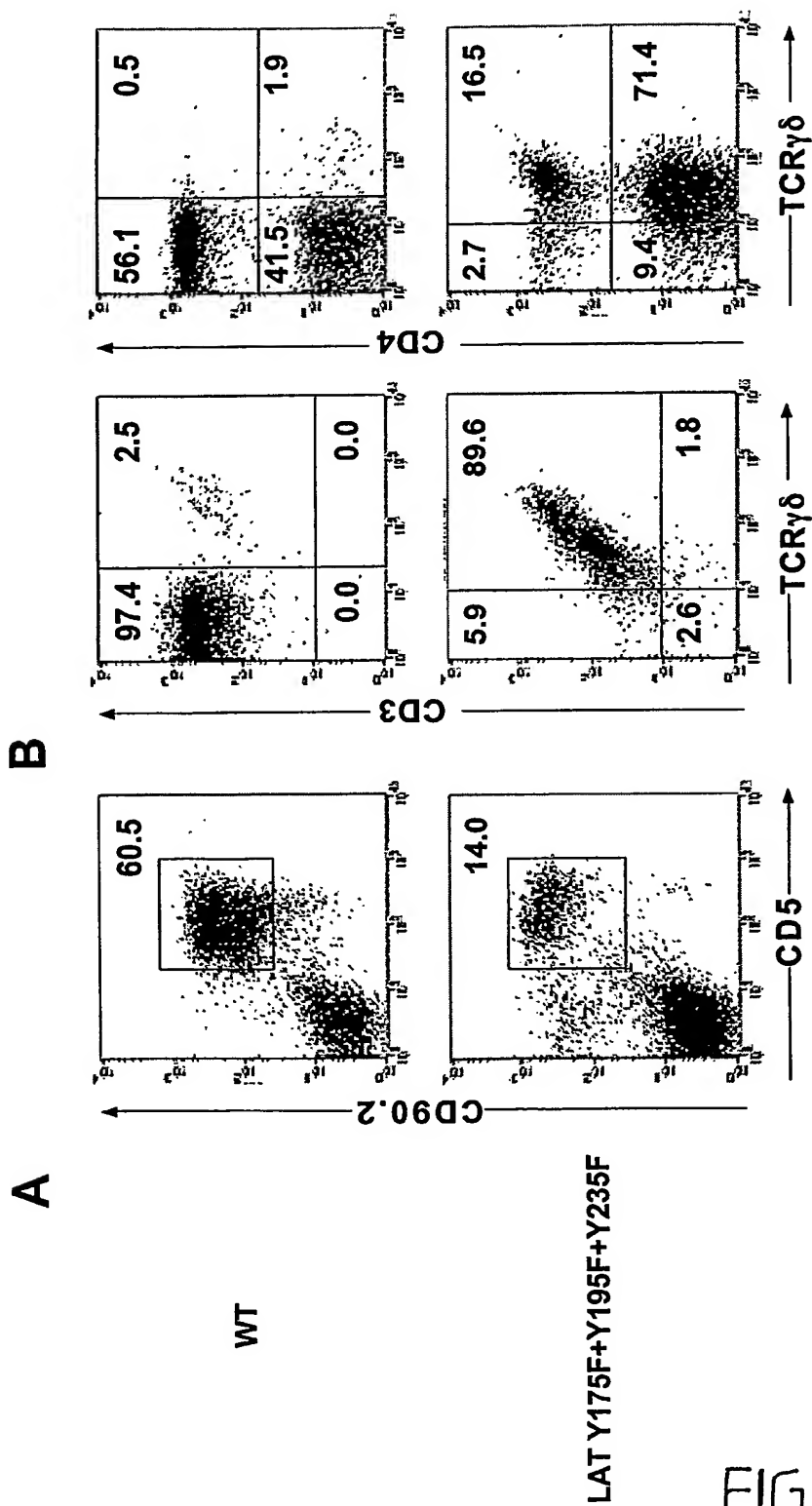


FIGURE 9

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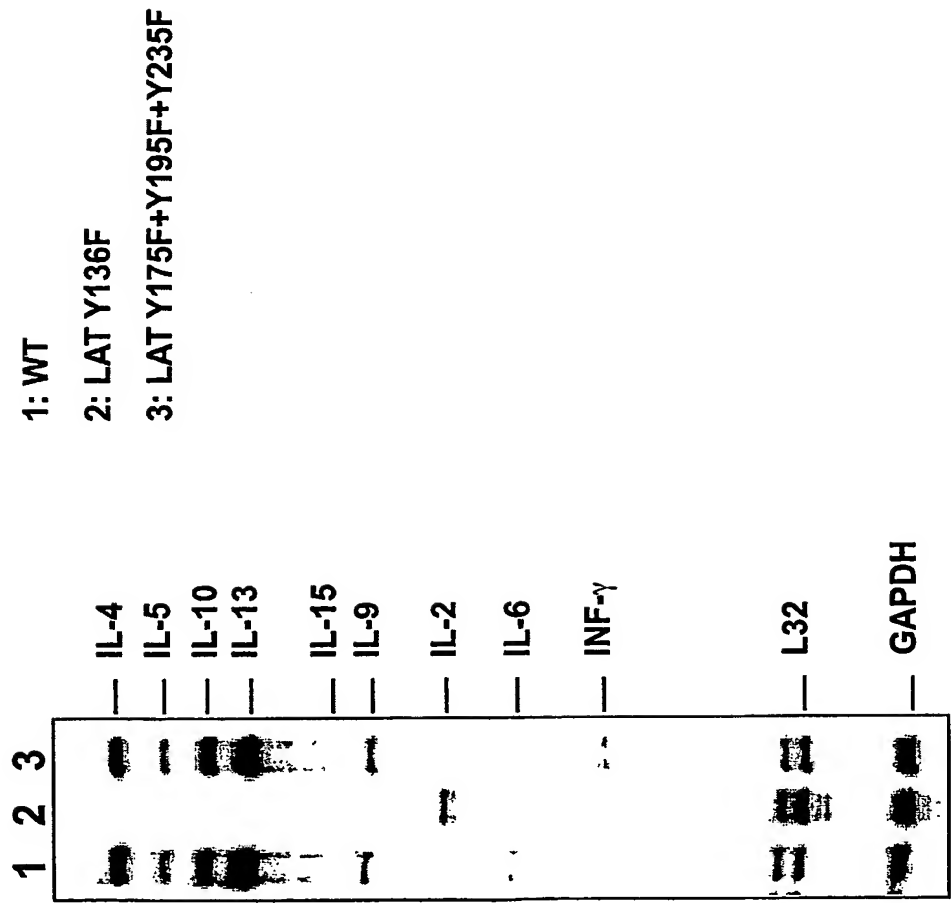


FIGURE 10

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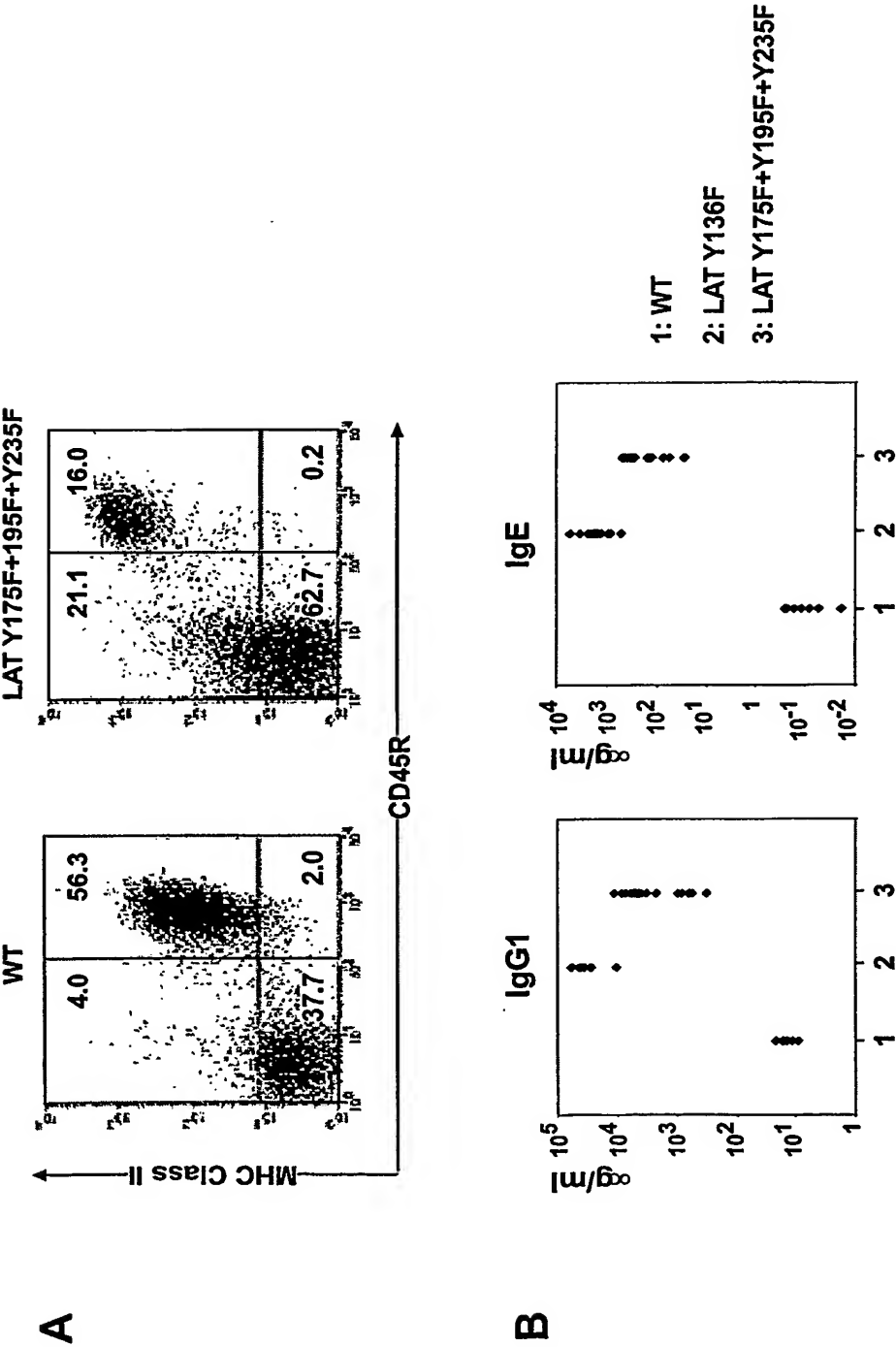


FIGURE 11

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21

(19) World Intellectual Property Organization
International Bureau



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21 August 2003 (21.08.2003)

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(10) International Publication Number
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15/63, C07K 14/47, A01K 67/027, C07K 16/18, G01N
33/68

(21) International Application Number: PCT/IB03/01044

(22) International Filing Date: 14 February 2003 (14.02.2003)

(25) Filing Language: English

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02290610.1 11 March 2002 (11.03.2002) EP

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(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Declaration under Rule 4.17:

— of inventorship (Rule 4.17(iv)) for US only

Published:

— with international search report

(88) Date of publication of the international search report:
13 November 2003

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: **MUTATED GENE CODING FOR A LAT PROTEIN AND THE BIOLOGICAL APPLICATIONS THEREOF**

(57) Abstract: The present invention relates to a mutated gene coding for a mutant LAT protein leading to an exaggerated T_H2 differentiation. The invention relates to biological structures containing said mutant, particularly, non-human LAT gene mutated animals, plasmids, chromosomal DNA's, embryos comprising said mutated gene, and applications thereof. The invention further relates to screening method for drug useful for treatment against asthma and allergy. Otherwise, the invention relates to method for producing IgE antibodies.



WO 03/068968 A3

INTERNATIONAL SEARCH REPORT

International Application No

PCT/IB 03/01044

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/12 C12N15/63 C07K14/47 A01K67/027 C07K16/18
G01N33/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K A01K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EMBL, SEQUENCE SEARCH, EPO-Internal, WPI Data, PAJ, BIOSIS, MEDLINE, CHEM ABS Dat

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	SOMMERS CONNIE L ET AL: "Knock-in mutation of the distal four tyrosines of linker for activation of T cells blocks murine T cell development." JOURNAL OF EXPERIMENTAL MEDICINE, vol. 194, no. 2, 16 July 2001 (2001-07-16), pages 135-142, XP002207177 ISSN: 0022-1007 cited in the application the whole document --- -/--	1,5-8, 11-13, 15,22



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

* Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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- *Z* document member of the same patent family

Date of the actual completion of the international search

30 July 2003

Date of mailing of the international search report

13/08/2003

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
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Fax: (+31-70) 340-3016

Authorized officer

Devijver, K

INTERNATIONAL SEARCH REPORT

Inter- national Application No
PCT/IB 03/01044

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	<p>AGUADO ENRIQUE ET AL: "Induction of T helper type 2 immunity by a point mutation in the LAT adaptor." SCIENCE (WASHINGTON D C), vol. 296, no. 5575, 2002, pages 2036-2040, XP002207182 14 June, 2002 ISSN: 0036-8075 the whole document -& DATABASE EMBL 'Online! 2 May 2002 (2002-05-02) MALISSEN M: "Mus musculus LAT gene for linker protein, exons 1-12." Database accession no. AJ438435 XP002207184</p>	1-34
P,X	<p>SOMMERS CONNIE L ET AL: "A LAT mutation that inhibits T cell development yet induces lymphoproliferation." SCIENCE (WASHINGTON D C), vol. 296, no. 5575, 2002, pages 2040-2043, XP002249670 14 June, 2002 ISSN: 0036-8075 the whole document</p>	1-34
A	<p>SAMELSON L E ET AL: "STUDIES ON THE ADAPTER MOLECULE LAT" COLD SPRING HARBOR SYMPOSIA ON QUANTITATIVE BIOLOGY, BIOLOGICAL LABORATORY, COLD SPRING HARBOR, NY, US, no. 64, 1999, pages 259-263, XP001056468 ISSN: 0091-7451 cited in the application page 261</p>	1-34
A	<p>LIN JOSEPH ET AL: "Identification of the minimal tyrosine residues required for linker for activation of T cell function." JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 276, no. 31, 3 August 2001 (2001-08-03), pages 29588-29595, XP002207178 ISSN: 0021-9258 cited in the application the whole document</p>	1-34
	-/--	

INTERNATIONAL SEARCH REPORT

Inte
nal Application No
PCT/IB 03/01044

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	ZHANG WEIGUO ET AL: "Association of Grb2, Gads, and phospholipase C-gamma1 with phosphorylated LAT tyrosine residues. Effect of LAT tyrosine mutations on T cell antigen receptor-mediated signaling." JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 275, no. 30, 28 July 2000 (2000-07-28), pages 23355-23361, XP002207179 ISSN: 0021-9258 cited in the application the whole document	1-34
A	WO 99 32627 A (SAMELSON LAWRENCE E ;US HEALTH (US); ZHANG WEIGUO (US)) 1 July 1999 (1999-07-01) * SEQ ID NO:3; SEQ ID NO:5 * claims 1-25	1-34
A	ZHANG WEIGUO ET AL: "Essential role of LAT in T cell development." IMMUNITY, vol. 10, no. 3, March 1999 (1999-03), pages 323-332, XP002207180 ISSN: 1074-7613	
A	SAITOH SHINICHIROH ET AL: "LAT is essential for FcepsilonRI-mediated mast cell activation." IMMUNITY, vol. 12, no. 5, May 2000 (2000-05), pages 525-535, XP002207181 ISSN: 1074-7613	

INTERNATIONAL SEARCH REPORT

International application No.
PCT/IB 03/01044

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Although claim 32 is directed to a diagnostic method practised on the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☒ Claims Nos.: 21
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 21

Claim 21 refers to a compound identified using a screening method according to any of claims 16 to 20 without giving a true technical characterization. Moreover, no such compounds are defined in the application. In consequence, the scope of said claim is ambiguous and vague, and its subject-matter is not sufficiently disclosed and supported (Art. 5 and 6 PCT). No meaningful search can be carried out for such a purely speculative claim whose wording is, in fact, a mere recitation of the result to be achieved.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Inter International Application No

PCT/IB 03/01044

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
WO 9932627	A	01-07-1999	AU 750543 B2	18-07-2002
			AU 2204799 A	12-07-1999
			CA 2316769 A1	01-07-1999
			EP 1141281 A2	10-10-2001
			WO 9932627 A2	01-07-1999

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